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## BCMB 406B Laboratory Schedule, Summer 2016

Week	Dates	
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## Evaluation and Assessment

### Percentage Breakdown for the Course:

**Lab Reports (25%):** Marks for the lab reports will be assigned as indicated below.

	<b>Marks</b>
Lab 1 – Primer Design	15
Lab 2 – Site-Directed Mutagenesis of a CBM Protein	45
Lab 3 – Purification and Characterization of a Mutant CBM Protein	40
<b>Total Marks</b>	<b>100</b>

**Lab Exams (50%):**

There will be three midterms in this course. Each midterm will be two hours in length and will only cover material from a single lab.

<b>Midterm</b>	<b>Date</b>	<b>Lab covered</b>	<b>Percent</b>
1	Tues. July 19	Lab 1	10 %
2	Tues. Aug. 2	Lab 2	20 %



## **Course Policies**

### ***Attendance***







16. **Dispose of infectious solid waste in the yellow biohazard buckets for autoclaving.**  
This includes pipette tips, agar plates, contaminated gloves or paper towels, etc
17. **Report any accidents or safety concerns to an instructor immediately.**
  - If skin comes into contact with chemicals, wash immediately with cold running water for at least 10 min.
  - In the event of a bacterial spill, pour an equal volume of bench disinfectant on top of the spill and allow it to sit for five minutes. Clean up the spill wearing gloves and using a no-touch technique. Discard all waste in a yellow biohazard bucket for autoclaving. **WASH YOUR HANDS with hand disinfectant and soap.**
  - Do not pick up broken glass. The instructor will do this.
  - If something has splashed in your eyes, rinse them at the eye wash station for at least 20 min.
18. **Mouth pipetting is prohibited.** A safety bulb or pipettor must be used.
19. **Sitting on laboratory work surfaces is prohibited.**
20. **Use of cellular devices is not allowed in the lab.**
21. **Note the location of the following safety equipment:**
  - Eye wash station
  - Safety shower
  - Fire extinguisher
  - Telephone
  - Fire alarm
22. **Before leaving the laboratory:**
  - Place all cultures and other contaminated materials to be discarded in the appropriate containers for sterilization in the autoclave
  - Put your experimental materials (**labeled!**) in the appropriate bins or racks for incubation or storage
  - Place contents of “tip discard” and used microfuge tubes into yellow biohazard bucket
  - Rinse all glassware and place in the appropriate bin(s)
  - Check that gas, water and microscope lights are turned off
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